

Standard Strategy for Comparison of the Nitrogen Fixation Rate in Different Samples

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Background & Objectives: Azotobacter, a gamma-proteobacterium, is an obligate aerobic free-living gram-negative soil bacterium capable of fixing nitrogen. There are several Methods for quantitative assay of nitrogen fixation. The discovery that the nitrogenase enzyme responsible for N₂-fixation also reduced C₂H₂ to C₂H₄ provided the Acetylene Reduction Assay (ARA) for the quantification of the N₂-fixation process. ARA is simpler and faster than the other Methods and it provides a highly sensitive and inexpensive way to quantify nitrogenase activity in N₂-fixing samples. This study aims at designing a standard strategy for comparison of the rates of N₂ fixation in different samples.

Methods: Five ml of the Azotobacter broth medium in 12 ml vials was inoculated with ~104CFU/ml of each isolate and incubated for 48-96 h at 28 °C. Once visible growth was observed, each vial was sealed with rubber stopper. By means of a disposable plastic syringe, 10% of air from the head space (7 ml) was removed and an equal amount of acetylene was injected into vials. Gas samples (0.7 ml) were removed after 24 h incubation, and were assayed for ethylene production with a gas chromatograph in triplicate. The rate of nitrogen fixation was calculated and values were obtained nmoles C₂H₄ h⁻¹ vial⁻¹.

Results: Among 8 isolates, only six were found to be able to fix atmospheric N₂. Amounts of acetylene reduced by isolates in samples were quite different. Rates obtained in isolates were in the range of 12.1 to 326.4 nmol C₂H₄ h⁻¹ vial⁻¹ while cell numbers were adjusted to 10⁷ CFU/ml.

Conclusion: Due to probable association between bacterial cell numbers and nitrogenase activity, we should estimate cell numbers of samples or final concentration of bacterial suspension adjusted according to CFU/ml during the course of ARA simultaneously.

Keywords: Azotobacter; Nitrogenase; Nitrogen Fixation; Acetylene Reduction Assay