

produce hydrogen peroxide upon the reaction by GOD. Hydrogen peroxide is produced gradually and does its antimicrobial activity in effective and gentle manner. GOD also is routinely used to evaluate glucose in physiological fluid included with medical diagnosing kits. Measurement of glucose concentration is important for determination of blood glucose levels in diabetic patients. GOD has been successfully immobilized onto various scaffolds to improve its stability. In this study we used covalent binding strategy for immobilization of GOD on glycation induced biocompatible amyloid nanofibers of bovine serum albumin in which glutaraldehyde was used as a cross-linker. The optimum concentration of the enzyme for immobilization was determined at 160 µg GOD per mg amyloid nano-fibers. The kinetic parameters of the free and immobilized GOD were also determined. Despite a decreasing of the catalytic performance (k_{cat}/K_m) of the enzyme upon immobilization, the covalently bound GOD on amyloid nano-fibers retained almost 40-70% of its activity after incubation at temperatures 25, 45 and 65°C but a major decrease in the activity was occurred for free enzyme under the same conditions. Hence, presented activity in a broad range of temperature most importantly at 40°C < in immobilized form compared to the free enzyme are explained. Interestingly, comparison between pH profiles of the free and immobilized enzymes indicates on increasing pH sensitivity of the GOD activity in the pH range lower than 6 and the optimum pH shift from 5 to 6 upon immobilization was observed.

Keywords: Catalytic Nano-Fibers, Amyloid, Glucose oxidase, Glycation, Immobilization.

Abstract No.208

Salt-induced Bundle Formation of F-actin Using a Detailed Model

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Actin filaments assemble into networks or bundles helped by linker proteins, depending on their different roles in cell function. Similar patterns have also been observed experimentally in solutions of F-actin rods with multivalent salt. In bundle structure, F-actins place close to each other in parallel formation. So despite their same high negative charge, why do they attract each other? Here we aim to represent a

model which contains enough structural details of F-actins to study the mechanism of this attraction. For this purpose, we apply the 4-sphere model for the structure of G-actin. In this model, G-actin is composed of four subdomains, one of which carries the most charge of G-actin (sd1). Using this 4-sphere model, we consider real structure and size of F-actin in addition to bending and twist rigidity and helical charge distribution of F-actins. Applying MD simulation to a group of these F-actins, we observe that they attract each and form a hexagonal lattice of the same lattice size as the experiment results. Also it is observed at equilibrium that counter-ions tend to assemble midline between two neighbor F-actins. This distribution of counter-ions around F-actins and how sd1s arrange on F-actins will shed light on understanding the mechanism. Accurate look at equilibrium details of arrangement of sd1 of two neighbor F-actins and plotting the force on them along F-actins, it is concluded that at equilibrium, F-actins will rotate and twist until they expose their sd1s close to each other. Because of their high negative charge, they will attract a cloud of counter-ions in a region between themselves. This cationic cloud will locally attract the two F-actins in its side and it is effectively seen that the two F-actins attract each other.

Keywords: F-actin, Bundle, Counter-ion, Multivalent salt, 4-sphere model, MD simulation.

Abstract No.209

Amyloid Peptide Nanofibrils: from Structure to their Lipid Peroxidative Effects

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Alzheimer's disease is a neurodegenerative disorder which is the most common cause of senile dementia. Amyloid beta (A β) is an amphiphilic peptide of 39 to 43 amino acids which is produced from a transmembrane precursor by a proteolytic cleavage, furthermore it is the main component of the neuritic plaques of the Alzheimer's disease. In this research some of the structural, physicochemical, and cytotoxic properties of the A β (25-35) and one of its modified, cysteine amyloid beta (25-35), which has an additional cysteine residue on its N-terminal, have been investigated. The objective of this research is to